

## Calculated Three Dimensional Structures of the Fumonisin B<sub>1-4</sub> Mycotoxins

R. C. Beier, M. H. Elissalde, L. H. Stanker

Food Animal Protection Research Laboratory, Agricultural Research Service,  
U.S. Department of Agriculture, 2881 F & B Road, College Station, Texas  
77845-9594, USA

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*Fusarium moniliforme* Sheldon is one of the most important ear rot pathogens of maize (*Zea mays* L.) and other grains throughout the world (Booth 1971). When fed to horses, corn infected with *F. moniliforme* produced clinical signs and lesions typical of the fatal neurotoxic syndrome, equine leukoencephalomalacia (ELEM) (Wilson and Maronpot 1971; Wilson *et al.* 1973). ELEM of horses and other equines is characterized by liquefaction necrosis of the white matter of one or both cerebral hemispheres (Wilson *et al.* 1985). ELEM is a disease reported in the U.S. since the early 1900's and known in Egypt, Africa, China, Japan, and European countries (Buck *et al.* 1979; Haliburton and Buck 1986; Marasas *et al.* 1988; McCue 1989). Corn samples associated with ELEM caused hepatotoxicity and renal toxicity in male Sprague-Dawley rats (Voss *et al.* 1989). *F. moniliforme* cultures caused porcine pulmonary edema (PPE) (Kriek *et al.* 1981; Ross *et al.* 1990), acute nephrosis and hepatitis in sheep, and cirrhosis, intraventricular cardiac thrombosis, and nephrosis in rats (Kriek *et al.* 1981), and were shown to have cancer-promoting activity (Gelderblom *et al.* 1988).

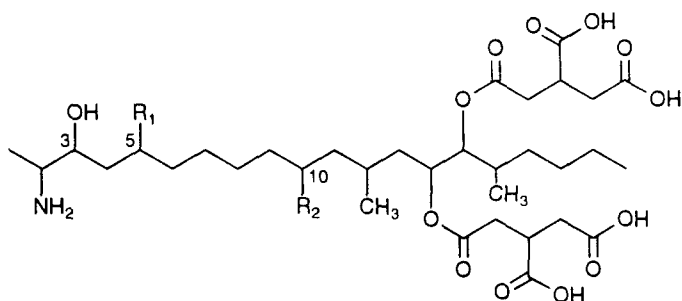
Four biologically active fumonisins have been isolated and characterized. Fumonisin B<sub>1</sub> and B<sub>2</sub> (FB<sub>1</sub> and FB<sub>2</sub>) (Fig. 1) (Bezuidenhout *et al.* 1988), and fumonisins B<sub>3</sub> and B<sub>4</sub> (FB<sub>3</sub> and FB<sub>4</sub>) (Fig. 1) (Gelderblom *et al.* 1992; Plattner *et al.* 1992) were isolated from the culture material of *F. moniliforme*. The first experimental evidence that FB<sub>1</sub> caused ELEM was presented by Marasas *et al.* (1988). Horses orally dosed with FB<sub>1</sub> developed ELEM (Kellerman *et al.* 1990), and ponies developed ELEM, hepatic necrosis, and encephalopathy (Ross *et al.* 1993).

In swine, PPE syndrome can be caused by IV injection of FB<sub>1</sub> (Harrison *et al.* 1990). From a series of studies with pigs, either PPE or liver failure emerged as

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Correspondence to: R. C. Beier



Fumonisin B<sub>1</sub>: R<sub>1</sub> = OH, R<sub>2</sub> = OH

Fumonisin B<sub>2</sub>: R<sub>1</sub> = OH, R<sub>2</sub> = H

Fumonisin B<sub>3</sub>: R<sub>1</sub> = H, R<sub>2</sub> = OH

Fumonisin B<sub>4</sub>: R<sub>1</sub> = H, R<sub>2</sub> = H

**Figure 1.** Chemical structures of the fumonisins calculated in this study.

distinct pathogenetic expressions of FB<sub>1</sub> toxicoses (Colvin *et al.* 1993). However, young female pigs fed a diet containing FB<sub>1</sub> developed nodular hyperplasia of the liver, and hyperplastic plaques within the distal esophageal mucosa (Casteel *et al.* 1993). It was shown that FB<sub>1</sub> undergoes enterohepatic cycling in the pig, and has affinity for specific tissues, particularly the liver (Prelusky *et al.* 1994).

Male and female rats fed diets fortified with FB<sub>1</sub> caused hepatotoxicity and cortical nephrosis in both sexes (Voss *et al.* 1993). Both the hepatic and renal lesions were consistent with those found in rats consuming *F. moniliforme*-infected corn (Voss *et al.* 1989; Plattner *et al.* 1990). Purified FB<sub>1</sub> is capable of inducing the subchronic liver and kidney lesions attributed to *F. moniliforme* cultures (Voss *et al.* 1993). FB<sub>1</sub> caused prenatal deaths and resorptions in hamsters (Floss *et al.* 1994). Turkey poult treated with FB<sub>1</sub> developed biliary hyperplasia, hypertrophy of Kupffer's cells, and thymic cortical atrophy (Weibking *et al.* 1993).

An isolate of *F. moniliforme*, strain MRC 826, was obtained from corn collected in Transkei, Union of South Africa, where the human esophageal cancer rate is high (Marasas *et al.* 1979). MRC 826 was found to be mutagenic (Gelderblom *et al.* 1983). The main fumonisin produced by *F. moniliforme*, FB<sub>1</sub>, was shown to have cancer-promoting activity similar to the fungal culture material (Gelderblom *et al.* 1988). A mammalian cell line of MDCK dog kidney epithelial cells was sensitive to FB<sub>1</sub> and FB<sub>2</sub> (Fig. 1) (Shier *et al.* 1991). FB<sub>2</sub> inhibited renal epithelial cell (LLC-PK) proliferation resulting in cell death (Yoo *et al.* 1992).

Biochemical findings show that FB<sub>1</sub> and FB<sub>2</sub> are the first naturally occurring inhibitors of sphingosine and sphinganine *N*-acyltransferase in rat primary hepatocytes (Wang *et al.* 1991). Fumonisin inhibit the conversion of sphinganine to *N*-acyl-sphinganine. It was suggested that the disruption of the *de novo* pathway of sphingolipid biosynthesis may be a critical event in the diseases associated with fumonisin consumption (Wang *et al.* 1991).

The fumonisins are a good example of a naturally occurring world wide environmental contamination caused by a fungus. FB<sub>1-3</sub> are the major naturally produced fumonisins from *F. moniliforme*, and FB<sub>4</sub> is produced in only minor quantities (Cawood *et al.* 1991). FB<sub>1</sub> is the most abundant member of the fumonisin family and usually accounts for about 70% of the total fumonisins (Plattner *et al.* 1992). As seen above, there are a large number of effects observed in animals and humans caused by FB<sub>1</sub>. There may be multiple effects in any one species, and the effects generally vary from specie to specie. However, there is only one known biochemical action of FB<sub>1</sub> and FB<sub>2</sub>; the inhibition of sphingolipid biosynthesis.

The purpose of this study is to determine the lowest energy 3-dimensional conformations available to FB<sub>1-4</sub> through molecular modeling methods. These structures may produce additional insight into other potential activities that may be associated with these molecules.

## MATERIALS AND METHODS

Molecular modeling studies were performed using a CAChe™ WorkSystem with release 3.5 software running on a Macintosh Quadra 700 equipped with a RP88 coprocessor board and a CAChe™ stereoscopic display (CAChe™ Scientific, Inc., Beaverton, OR). Minimum potential energy conformations of all compounds shown in Fig. 2 and 3 were calculated using molecular mechanics with Allinger's standard MM2 force field parameters (Allinger 1977) augmented to contain force field parameters for cases not addressed by MM2 (CAChe™ Scientific, Inc.). A good description of molecular mechanics and its underlying philosophy is presented by Boyd and Lipkowitz (1982). Molecular mechanics calculations utilized the conjugate gradient optimization technique. Following the initial geometrical optimization, a one pass sequential search to determine the lowest potential energy conformations was performed by rotating all dihedral angles through 360° in 15° increments. Following the initial sequential search, multiple multipass sequential searches of four dihedral angles through 360° in 15° increments at a time were conducted to determine the lowest potential energy for the interacting systems of atoms. One final multipass sequential search of each set of dihedral angles through 90° or 180° in 5° to 7.5° increments was conducted for refinement of the optimum geometries. At completion of each sequential search a geometrical optimization was performed on each of the 5 to 10 selected structures

having the lowest potential energies. The resultant lowest potential energy structure was then used for the next refinement calculation. The set of structures resulting from each sequential calculation were viewed using the CAChe™ Visualizer+ graphics application.

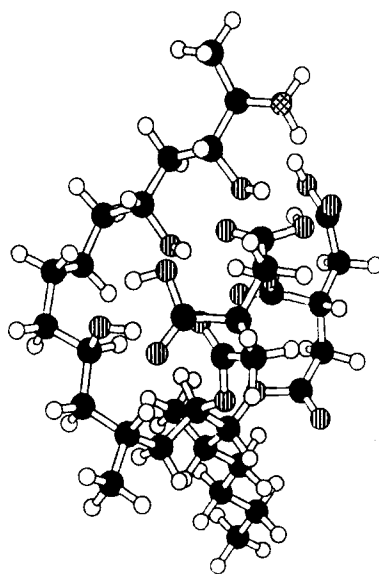
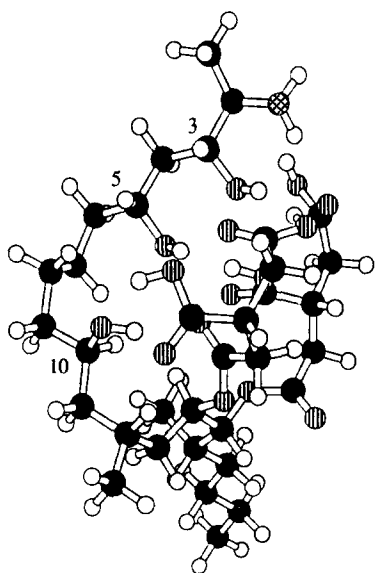
To obtain the electron density distribution of FB<sub>1</sub>, the final structure of FB<sub>1</sub> as obtained from molecular mechanics calculations was then re-calculated using ExtHückel to obtain all of the molecular orbitals. The resultant file was submitted to the tabulator to convert the results of the computational methods into 3D coordinates required by the CAChe Visualizer+ application. The final results showing the electron density of FB<sub>1</sub> was observed using the Visualizer+ application.

Stereo ball and cylinder representations were copied to the clipboard in the print preview mode. These figures were pasted into McDraw II for sizing and attaching coding on the spheres to represent the atom type. The ball and cylinder models were printed on a Macintosh Personal LaserWriter at 300 dots/inch and the electron density models were printed on a Macintosh LaserWriter Select 360 at 600 dots/inch using the following Macintosh print settings: high-quality shaded, color/gray scale, high resolution and faster postscript printing mode.

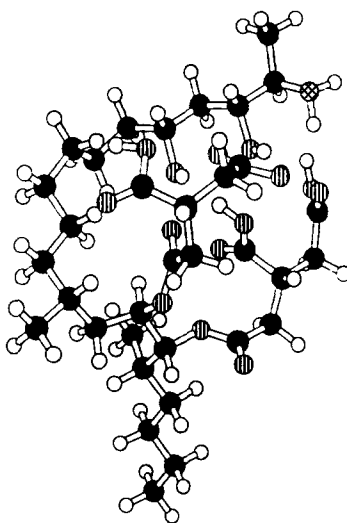
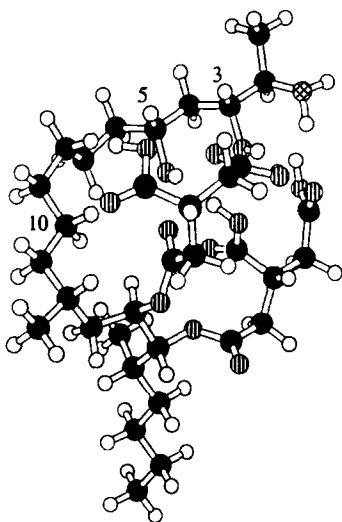
## RESULTS AND DISCUSSION

The molecular mechanics calculated structures for FB<sub>1</sub> and FB<sub>2</sub> (Fig. 2), and FB<sub>3</sub> and FB<sub>4</sub> (Fig. 3) are represented as ball and cylinder stereo models. Carbon atoms are depicted as black spheres, hydrogen atoms are open spheres, nitrogen atoms are cross-hatched spheres, and oxygen atoms are represented as spheres with vertical lines. The use of a stereoscope gives each set of molecules a three dimensional appearance. The four molecules each have a unique folded structure that one might liken to that of a folded peptide. FB<sub>1</sub> has more internal hydrogen bonding with the hydroxyl moiety at both C-5 and C-10 than does FB<sub>2</sub> or FB<sub>3</sub> which have only one hydroxyl at C-5 and C-10, respectively. FB<sub>4</sub> has the least potential internal hydrogen bonding with no hydroxyls at either the C-5 or C-10 positions.

A very interesting cage-like structure is formed by the folding of the amine backbone with the two esterified trimethylpropane-1,2,3-tricarboxylic acid side chains. The loss of a hydroxyl at C-5 or C-10 in FB<sub>2</sub> and FB<sub>3</sub>, respectively, allows this cage to become smaller than predicted for FB<sub>1</sub>. The three dimensional structure formed is similar to the claw of a bird. Recently, the observance of the fumonisins folding into this cage through molecular modeling has allowed the postulation of why the monoclonal antibodies produced to FB<sub>1</sub> linked through the amino group have lower specificities for free FB<sub>1</sub> than expected (Elissalde *et al.* 1995). This cage-like structure brings the amine, hydroxyls, and carboxylic acid groups into

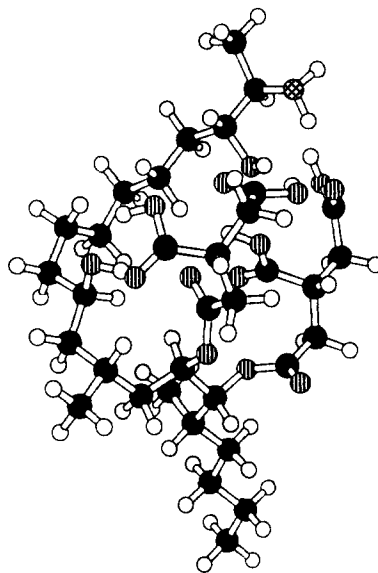
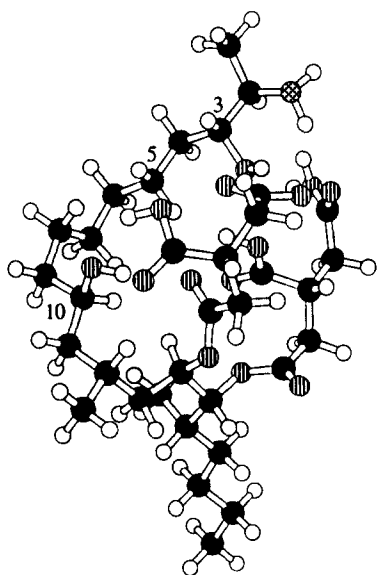


Fumonisin B<sub>1</sub>

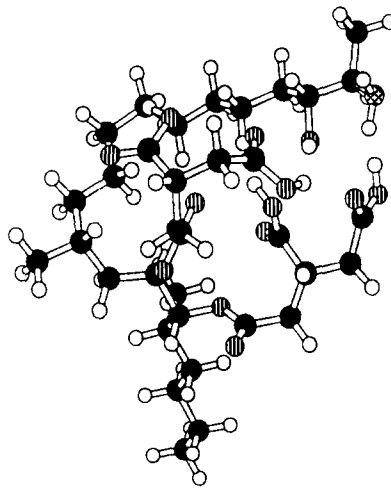
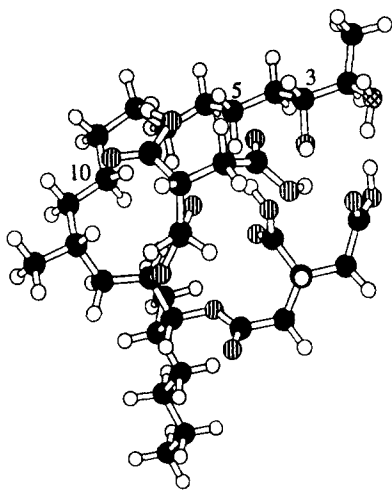


Fumonisin B<sub>2</sub>

**Figure 2.** Ball and cylinder stereo-models of the minimum energy conformations of fumonisins B<sub>1</sub> and B<sub>2</sub>. Carbon atoms are depicted as black spheres, hydrogen atoms are open spheres, nitrogen atoms are cross-hatched spheres, and oxygen atoms are represented as spheres with vertical lines.

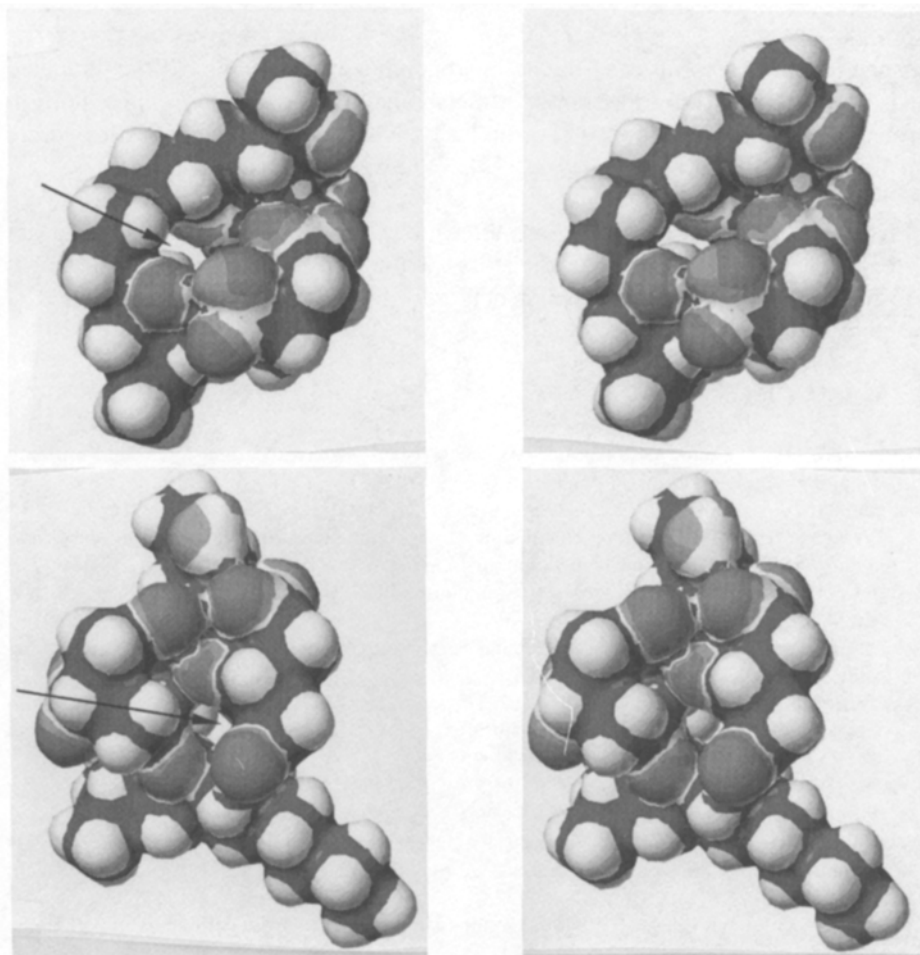


Fumonisin B<sub>3</sub>



Fumonisin B<sub>4</sub>

**Figure 3.** Ball and cylinder stereo-models of the minimum energy conformations of fumonisins B<sub>3</sub> and B<sub>4</sub>. Carbon atoms are depicted as black spheres, hydrogen atoms are open spheres, nitrogen atoms are cross hatched spheres, and oxygen atoms are represented as spheres with vertical lines.



**Figure 4.** Left and right stereo views of the electron density distribution for FB<sub>1</sub> looking at the electrophilic end (top view) and lipophilic end (bottom view). The arrows show spaces within FB<sub>1</sub> that traverse from the electrophilic to the lipophilic part of the molecule.

close proximity. Both the structures of these molecules and the close association of the amine, carboxylic acid groups, and hydroxyls suggests that these molecules may have chelator-type activity.

Fig. 4 shows two stereo views of the electron density distribution for FB<sub>1</sub>. The use of a stereoscope gives each set of stereo views a three dimensional appearance. The top view shows the electrophilic end of the molecule and the bottom view is a side presentation that shows the lipophilic end. In both views, arrows point out large channels within the center of FB<sub>1</sub> that have no electron density. This space or hole begins on the electrophilic end and transverses the molecule, bifurcating, and opening in two locations to either side of the lipophilic end. These spaces may

change in size as a result of changing hydrogen bonding patterns. If the lipophilic end of FB<sub>1</sub> were imbedded into a membrane with the electrophilic portion protruding; the membrane may become leaky due to these spaces or holes within the fumonisin structure.

Based upon structural interpretations after molecular modeling of FB<sub>1</sub>, it appears that these molecules may act as chelators, or they may cause membrane leakage. This hypothesis will require experimental work for corroboration of the structure reactivity relationship.

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